



## **TOOLS Water DNA & RNA Extraction Kit**

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## Introduction

TOOLS Water DNA & RNA Extraction Kit is a buffer-based system for the extraction and purification of DNA and RNA collected from filter paper. Simple centrifugation procedures enable the complete removal of contaminants and enzyme inhibitors. It is a fast, simple, and cost-effective method, and the purified DNA and RNA are suitable for downstream applications.

## Kit Contents

Contents	TX-WT01 (50 preps)
Buffer TWA	30 ml
Buffer TWB	12 ml
Buffer TWC	300 µl
Binding Gel	50 tubes

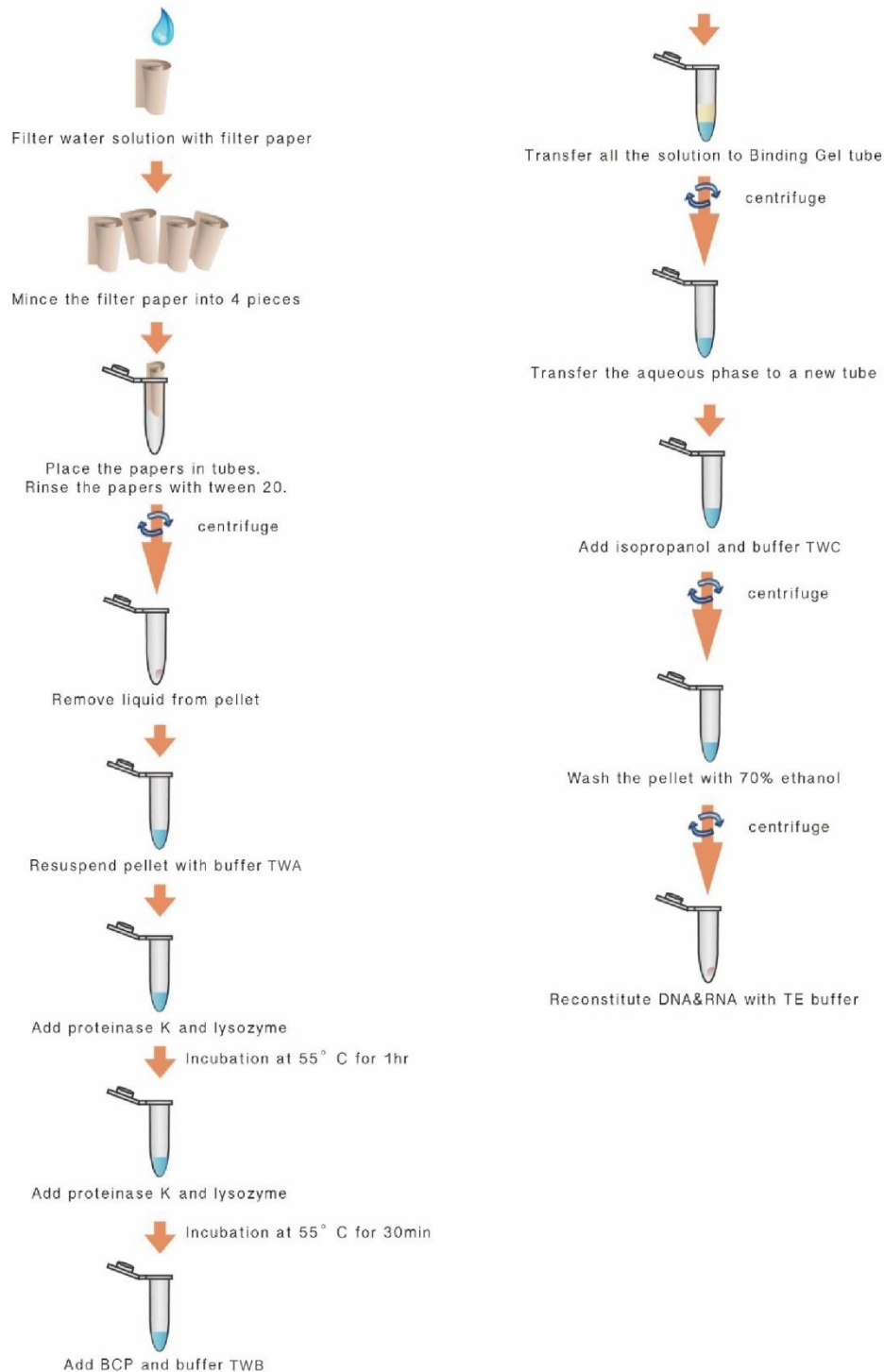
### Storage

TOOLS Water DNA & RNA Extraction Kit can be stored at room temperature for up to 24 months

### Materials not provided

- 1-Bromo-3-chloropropane (BCP; CAS Number: 109-70-6).
- TOOLS Proteinase K (Cat. No. RTT-BD03) or other compatible reagents and lysozyme.

## Workflow



## Protocol

### Sample preparation

Collect 500 mL of aqueous solution and centrifuge at 1,000 rpm for 5 min. Carefully collect 450 mL from the tube and filter the solution through 0.22- or 0.45- $\mu$ m filter paper (Millipore, Merck).

2. Cut the filter paper into four pieces and mince. Place the paper in a 50-mL tube (the air collecting side of the papers should face the center of the tube). Collect filtrate by rinsing the paper with 0.2% Tween-20 and shake the tube gently for 60 s to remove the filtrate from the paper.
3. Remove the paper from the tube, centrifuge at 14,000 rpm for 3 min, and remove as much liquid as possible.
4. Add 500  $\mu$ L of Buffer TWA to the tube and vortex for 60 s to resuspend the pellet thoroughly.
5. Add 5  $\mu$ L proteinase K (20 mg/mL) and 5  $\mu$ L of lysozyme (100 mg/mL) to the tube, vortex for 60 s, and incubate for 1 h at 55 °C. Vortex the tube for 20 s to mix thoroughly.

### DNA and RNA extraction

Add 5  $\mu$ L of proteinase K and 5  $\mu$ L of lysozyme to the tube. Vortex for 60 s and incubate at 55 °C for 30 min.

Add 200  $\mu$ L of BCP and 200  $\mu$ L of Buffer TWB to the tube and mix the sample by inverting the tube three times. Transfer all of the solution to the binding gel tube (centrifuge at  $13,000 \times g$ , 30 s before use).

3. Invert the binding gel tube three times (do not vortex) and centrifuge at  $12,000\text{--}16,000 \times g$ , 5 min.
4. Transfer supernatant to a new microcentrifuge tube and add 5  $\mu$ L of Buffer TWC and 550  $\mu$ L of isopropanol to the tube.
5. Invert the tube three times and incubate at 37 °C for 10 min.
6. Centrifuge 14,000 rpm for 5 min. Remove supernatant.
7. Add 600  $\mu$ L of 70% ethanol. Centrifuge 14,000 rpm for 5 min. Remove supernatant.
8. Air dry the pellet and rehydrate it with 20–50  $\mu$ L of TE buffer or ddH<sub>2</sub>O.  
(adjust buffer volume according to pellet size).

This product is for research only. Not for diagnostic or clinical use.